Instructions for Use

RealLine Trichomonas vaginalis
Str-Format

A QUALITATIVE ASSAY KIT FOR THE DETECTION OF TRICHOMONAS VAGINALIS DNA BY REAL TIME PCR METHOD

In vitro Diagnostics

CE

RealLine Trichomonas vaginalis (Str Format)	**VBD2098**	**96 Tests**
valid from	**September 2019**	

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Explanation of symbols used in labeling

IVD	*In vitro* diagnostic medical device
LOT	Batch code
REF	Catalogue number
	Contains sufficient for <n> tests
	Use-by-date
	Temperature limit
	Consult instructions for use
	Keep away from sunlight
	Manufacturer

BIORON Diagnostics GmbH

In den Rauhweiden 20
67354 Römerberg
Germany

Phone +49 6232 298 44 0
Fax: +49 6232 298 44 29
info@bioron.de

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ASSAY KIT FOR THE QUALITATIVE DETECTION OF *TRICHOMONAS VAGINALIS* DNA BY REAL TIME PCR METHOD
In vitro Diagnostics

1. INTENDED USE

Clinical information
Trichomonas vaginalis - a flagellate - causes Trichomoniasis, a disease that causes vaginal infections in women and urethritis in men. The transmission of Trichomoniasis runs through sexual intercourse and is one of the most common sexually transmitted diseases.

RealLine Trichomonas vaginalis (Str-Format) assay kit is designed to detect *Trichomonas vaginalis* DNA isolated from clinical specimens using extraction kits:
RealLine DNA-Express (REF VBC8899)
RealLine DNA-Extraction 2 (REF VBC8897)
RealLine DNA-Extraction 3 (REF VBC8889)
RealLine Extraction 100 (REF VBC8896)

RealLine Trichomonas vaginalis (Str-Format) kit is designed for the analysis of clinical materials: swabs of the epithelial cells, semen, prostatic fluid and urine.

The assay is based on Real Time polymerase chain reaction (PCR) method with fluorescent detection of amplified products.

The **Str-Format Kit** contains 96 tubes (0.2 ml) in strips with lyophilized Mastermix. 50 µl of extracted DNA have to be pipetted into the tube and the ready mastermix is diluted. The kit contains reagents required for 96 tests, including control samples and the positive control sample.

The kit is validated for use with block-type cyclers:iQ™5 iCycler, iQ™ iCycler, CFX™96 (Bio-Rad, USA), DT-96 (DNA-Technology, Russia) and RealLine Cyclers (BIORON Diagnostics GmbH)

The use of:

- **Extraction Kits for nucleic acids from clinical specimen from other supplier**
- **other real-time PCR devices**
- **appropriate reaction volumes, other than 50 µl**

has to be validated in the lab by the user. The special notes regarding the internal control IC have to be strongly followed.

2. KIT CONTENTS

Universal Positive Control Sample (PC)	1 vial, 1 ml
Ready Master Mix (RMM), lyophilized	96 test-tubes (12 strips x 8 tubes)
The kit is additionally supplied with optical-transparent PCR-film	

3. PRINCIPLE OF THE METHOD

Real time PCR is based on the detection of the fluorescence, produced by a reporter molecule, which increases as the reaction proceeds. Reporter molecule is dual-labeled DNA-probe, which specifically binds to the target region of pathogen DNA. Fluorescent signal increases due to the fluorescent dye and quencher separating by Taq DNA-polymerase exonuclease activity during amplification. PCR process consists of repeated cycles: temperature denaturation of DNA, primer annealing and complementary chain synthesis.

Threshold cycle value – Ct – is the cycle number at which the fluorescence generated within a reaction crosses the fluorescence threshold, a fluorescent signal rises significantly above the background fluorescence. Ct depends on initial quantity of pathogen DNA template.

The use of **Internal Control (IC)** prevents generation of false negative results associated with possible loss of DNA template during specimen preparation. IC indicates if PCR inhibitors occur in the reaction mixture. IC template should be added in each single sample (including control samples) prior to DNA extraction procedure. The amplification and detection of IC does not influence the sensitivity or specificity of the target DNA PCR.

Note: IC is a component of the NA extraction kits of RealLine series. Internal Control is added to the sample during NA isolation step and is used throughout the whole process of NA extraction, amplification, detection.

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4. SPECIFICATIONS

4.1. Sensitivity:

Sensitivity control was performed on samples in containing Standard Reference Samples containing 100 copies of *Trichomonas vaginalis* DNA per sample – equals 100%.

4.2. Specificity:

Specificity of *Trichomonas vaginalis* DNA detection using Standard Reference Panel of negative samples (DNA extracts) - equals 100%.

4.3. Diagnostic sensitivity:

Diagnostic sensitivity of *Trichomonas vaginalis* DNA detection: clinical trials conducted on 40 positive samples showed 100% sensitivity (interval 93% -100%, with a confidence level of 90%);

4.4. Diagnostic specificity

Diagnostic specificity of *Trichomonas vaginalis* DNA detection: clinical trials conducted on 40 negative samples showed 100% specificity (93% -100% interval, with a 90% confidence level).

Analysis by the CE-marked reference kit showed full match of results.

5. LIMITATIONS

- This assay must not be used on the clinical specimen directly. Appropriate nucleic acids extraction methods have to be conducted prior to using this assay.
- The presence of PCR inhibitors (e.g. heparin) may cause false negative or invalid results.
- When monitoring a patient the same extraction method must be used in all determinations. Otherwise, results may not be relative.
- The kit is designed for use in patients with a clinical history and/or symptoms consistent with Trichomonas infection. The kit may be used for screening purposes.
- Diagnostic sensitivity of the kit may vary depending on the pathogen prevalence and characteristics of the enrolled cohort.
- Reliable results depend on adequate specimen sampling.
- Positive results indicate active or asymptomatic infection; results should be interpreted with consideration of clinical and laboratory findings.
- Negative results indicate lack of detectable DNA but do not exclude the infection or disease.
- Potential mutations within the target regions of the *Trichomonas vaginalis* genome covered by the primers and/or probes used in the kit may result in failure to detect the presence of the pathogens.
- The kit is not intended to replace culture and other methods (e.g., cervical exam) for diagnosis of urogenital infection.

6. WARNING AND PRECAUTIONS

- For In vitro use only.
- The kits must be used by skilled personnel only.
- When handling the kit, follow the national safety requirements for working with pathogens.
- To prevent contamination, the stages of DNA isolation and PCR test run must be spatially separated.
- Avoid microbial and ribonuclease contamination of reagents when removing aliquots from reagent vials.
- Wear protective disposable gloves, laboratory coats and eye protection when handling specimens and kit reagents.
- Every workplace must be provided with its own set of variable-volume pipettes, necessary auxiliary materials and equipment. It is prohibited to relocate them to other workplaces.
- The use of sterile disposable pipette tips is recommended.
- Never use the same tips for different samples.
- Do not pool reagents from different lots or from different vials of the same lot.
- Dispose unused reagents and waste in accordance with country, federal, state and local regulations.
- Do not use the kit after the expiration date at the side label of the kit.

7. ADDITIONAL MATERIALS AND DEVICES REQUIRED BUT NOT SUPPLIED

- real time PCR system, like described in p.1
- DNA-Extraction Kit: RealLine DNA Express, RealLine DNA-Extraction 3, RealLine DNA-Extraction 2 or RealLine Extraction 100
- Internal Control reagent (VBC8881) and Negative Control Sample if the kit is used with the extraction kits of other supplier;
- laminar safety box;
- refrigerator;
- half-automatic variable-volume single-channel pipettes;
- disposable medical non-sterile powder-free gloves;
- disposable pipette tips with aerosol barrier;
- biohazard waste container.

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8. PREPARATION OF THE SPECIMEN

Each group of samples undergoing the procedure of DNA isolation must include a Positive Control sample (PC) from this kit and a Negative Control sample (NC) which is a component of the DNA extraction kit.

We strongly recommend the implementation of the Internal Control IC, the Negative Control NC and Positive Control PC samples to the extraction procedure.

If using a kit of another supplier for the extraction of nucleic acids as recommended in p1., add **20 µl** of **IC (VBC8881)** to each tube.

- For the NC use **100 µl** of the Negative Control Sample
- For the PC use **70 µl** of Negative Control Sample and **30 µl** of Positive Control to the tube marked PC.

Prepare the samples for the assay using **RealLine DNA-Express, RealLine DNA-Extraction 2, RealLine DNA-Extraction 3** or **RealLine Extraction 100** extraction kits according their instruction manuals.
If samples of isolated DNA were stored frozen prior the assay, thaw them and keep at least 30 minutes at a temperature of (18 - 25) °C.

The isolated DNA can be stored at (2 – 8) °C for 24 hours .
After initial opening shelf life of Positive Control sample (PC) at (2 – 8) °C is no more than 1 month or in 50 µl aliquots no more than 3 month at (-18…-24) °C.

9. PROCEDURE

9.1. Preparation of the reagents.

Prior the test take the kit out of the refrigerator and keep the **Ready Master Mix (RMM)** closed in the package at (18 – 25) °C for at least 30 minutes. Then open the package and cut the necessary number of tubes in strips with RMM (*including prepared samples and controls: 1 NC and 1 PC*) with the knife. The cutted tubes hold together with the covering film.
Put the remaining strips immediately back into the foil pouch, squeeze the air out and tightly close with the clip.
After initial opening the shelf life of RMM at (2 – 8) °C is 3 months.

9.2. Label the tubes with RMM for each specimen and control.
Attention! *Labels should be placed on the lateral side of the tubes.*

9.3. Add **50 µl** of corresponding isolated DNA solution to each tube using a separate pipette tip with filter. Do not touch the pellet! Tightly close the tubes with caps or seal with the PCR transparent film.

9.4. Place the tubes into the Real Time PCR system.

9.5. Program Real Time PCR system as follows:

Stage 1:	50°C	2min	
Stage 2:	95°C	2min	
Stage 3:	94°C	10 sec	50 cycles
	60°C*	20 sec	

* Measure the fluorescence at 60°C

9.6. Select the amplification detection channels:

- Collect Real Time PCR data through the **FAM** channel for detection of amplification of IC DNA.
- Collect Real Time PCR data through the **ROX** channel for detection of amplification of infectious agent's DNA.

9.7. Program the positions of test tubes with samples, positive and negative controls according to the instruction manual for the Real Time PCR system in use.

9.8. Run the program.

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10. DATA ANALYSIS AND INTERPRETATION

10.1 For **PC** the program should detect:

- increase of the IC DNA amplification signal (channel **FAM**) and determine the threshold cycle, IC **Ct**;
- increase of the *Trichomonas vaginalis* DNA amplification signal (channel **ROX**) and determine the **Ct** value.

10.2 For **NC** the program should detect the increase of the amplification signal of IC DNA (channel **FAM**) and determine the threshold cycle, IC **Ct**. No significant **ROX** fluorescent increase should appear (*no Trichomonas vaginalis DNA amplification*).

10.3 For each specimen the program should detect the increase of the amplification signal of IC DNA (channel **FAM**) and determine IC **Ct**.

10.4 Calculate (IC **Ct**)$_{av}$ as an average IC **Ct** of all analyzed samples (including PC and NC). IC **Ct** values that differ by more than 2 from the (IC **Ct**)$_{av}$ should be ignored. Recalculate the (IC **Ct**)$_{av}$ for the remaining values after the screening.

10.5 The sample is considered **negative** (not containing *Trichomonas vaginalis* DNA), if **Ct** value via **ROX** channel for this sample is **above 40** or is not determined.

When IC **Ct** value for such sample differs from the (IC **Ct**)$_{av}$ value by more than 2, the result is regarded as equivocal. A repeated analysis of the sample, starting with the DNA isolation step is necessary.

10.6 The sample is considered **positive**, i.e. contains *Trichomonas vaginalis DNA*, when **Ct** value via **ROX** channel for this sample is **less than or equals to 40.**

10.7 When Ct value for NC through **ROX** channel **is less than or equal to 40**, this indicates the presence of contamination. In this case all positive results of this individual PCR run are considered as equivocal. Actions are required to identify and eliminate the source of contamination, and repeat the analysis of all samples of this run that were identified as positive. Samples that showed negative results in this run should be considered as negative.

11. STORAGE AND TRANSPORTATION

- Store and transport the assay kit at (2 - 8) °C in the manufacturer's packing.
- Transport at (2 - 8) °C . Transportation at up to 25 °C for up to 10 days is allowed.
- Do not freeze the kit!
- Do not pool reagents from different lots or from different vials of the same lot.
- Strictly follow the Instruction manual for reliable results.
- Do not use kits with damaged inner packages and get in contact with BIORON Diagnostics GmbH.

- **Storage and shelf life of solutions and components of the kit after initial opening:**
  Positive Control sample: 1 month at (2 - 8) °C or in 50 µl aliquots at minus (18 - 60) °C for up to 3 months.
  Ready Master Mix (RMM): 3 months at (2 – 8) °C

Technical Support: techsupport@bioron.de

ANNEX I: Settings for RealLine Cycler and DT96:
for these cyclers the measurement exposure must be adjusted. Choose the **Operation with the device** mode in the **Settings** menu, select the item **Measurement exposition:**

- **FAM** to **250**
- **HEX** and **ROX** to **1000**

Confirm that the current exposure value is saved by pressing **YES**
Attention! The specified exposure values are applicable only for RealLine kits and, if necessary, must be changed for other purposes.

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BIORON Diagnostics GmbH
info@bioron.de - www.bioron.de